

Research Symposium

Pseudogenization of a resistance-nodulation-division (RND) efflux pump subunit underlies drug and cell envelope stress sensitivity in *Brucella ovis*

Thomas Kim¹, Bill Hong¹, Erika Lisabeth¹, Richard Neubig¹, Aretha Fiebig¹, Sean Crosson¹

¹ College of Osteopathic Medicine, Michigan State University

<https://doi.org/10.51894/001c.143477>

Spartan Medical Research Journal

Vol. 10, Issue 2, 2025

01

Brucella species are intracellular bacterial pathogens that pose a significant threat to both animal and human health. To identify compounds that reduce *Brucella ovis* fitness in mammalian phagocytes, we conducted a high-throughput luminescence-based screen, which revealed dihydropyridine-class Ca²⁺ channel blockers nicardipine and cilnidipine as potential host-targeting anti-infectives. However, follow-up dose-response studies in pure culture revealed that these compounds directly inhibit *B. ovis* growth. To investigate possible dihydropyridine resistance mechanisms, we selected for *B. ovis* mutants tolerant to these drugs and identified single-base deletions in the *bepE* pseudogene. These mutations restored a functional open reading frame for BepE, a subunit of a resistance-nodulation-division (RND) efflux pump, increasing *B. ovis* resistance to dihydropyridine treatment. Given that *B. ovis* has undergone extensive pseudogenization and exhibits greater chemical

susceptibility than other *Brucella* species, we examined whether *bepE* influenced cell envelope integrity. *B. ovis* mutants with an intact *bepE* gene displayed enhanced resistance to membrane disruptors, including deoxycholate. To extend these findings, we investigated *bepE* function in *Brucella abortus*, a closely related zoonotic pathogen that encodes a fully intact BepE protein. Deleting *bepE* in *B. abortus* increased its susceptibility to deoxycholate and its sensitivity to cilnidipine during macrophage infection, indicating that *bepE* not only contributes to drug resistance in the intracellular niche but also supports *B. abortus* resistance to cell envelope stress. The results define *bepE* as a determinant of *Brucella* resistance to antimicrobial compounds, and demonstrate that its pseudogenization contribute to the heightened chemical sensitivity of *B. ovis* relative to other classical *Brucella* species.

Published: September 30, 2025 EST.



This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CCBY-4.0). View this license's legal deed at <http://creativecommons.org/licenses/by/4.0> and legal code at <http://creativecommons.org/licenses/by/4.0/legalcode> for more information.